Adenosine and AMP Gene Expression Profiles Predict Response to Adenosine Pathway Therapies and Indicate a Need for Dual Blockade of CD73 and A2AR with CD73 Inhibitors

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ADENOSINE INHIBITS ANTI-TUMOR IMMUNITY



- Extracellular adenosine has a short half life and it is not feasible to routinely measure in human tumors
- Determine genes/proteins modulated by adenosine as a surrogate to identify patients with adenosine rich tumors

ADENOSINE INDUCES A SPECIFIC GENE SIGNATURE

HUMAN PBMCs STIMULATED WITH NECA, A STABLE ADENOSINE ANALOG

Experiment	Human	+ NECA		+ ACTIVATION		NanoString
Setup	PBMC			anti-CD3/CD28		
			1 hour		48 hours	

ADENOSINE SIGNATURE - NANOSTRING

	Adjusted p Value	Function
IL23A	1.44E-04	Increases angiogenesis and reduces CD8+ T-cell infiltration
SLC11A1	1.27E-03	Natural resistance-associated macrophage protein 1
CXCL2	1.27E-03	MIP2a: macrophage inflammatory protein 2, alpha
CXCL7	1.27E-03	PPBP. Pro-Platelet Basic Protein
CXCL6	1.40E-03	GCP2: Granulocyte chemotactic protein 2
CXCL3	1.40E-03	Controls migration and adhesion of monocytes
IL-6	1.48E-03	Pro- and anti-inflammatory cytokine
IL-1a	1.73E-03	Inflammation
CXCL8	1.98E-03	IL-8. Nutrophil chemotactic factor
CXCL5	1.98E-03	Attracts and activates neutrophils
THBS1	2.28E-03	Multiple functions. Inhibits angiogenesis & immune regulation
IL-1β	2.38E-03	Inflammation
PTGS2	2.65E-03	COX-2. Elevated during inflammation and cancer
IL-24	2.70E-03	Cell survival and proliferation. Activates STAT1/3
CXCL1	3.92E-03	Neutrophil chemotractant
CD86	5.31E-03	B7-2: Costimulatory signal for T cell activation and survival
CLEC5A	5.82E-03	Interacts with DAP-12 and may play a role in cell activation
CD14	6.24E-03	Expressed by myeloid cells

Human PBMCS were isolated from buffy coat samples by density centrifugation with Histopaque 1077 (400*g, 30 min). Cells were washed and resuspended at a density 2*10⁶ cells/ml in RPMI + 10% human serum. PBMCs were stimulated with

DMSO or 5'-N-Ethylcarboxamidoadenosine (NECA) at 0.1, 1, or 10 µM for one hour. T cells were then activated with anti-CD3 + anti-CD28 antibodies and incubated for 48 hrs. Purified RNA was collected using a Qiagen RNAEasy Kit and gene

expression analysis was performed using the NanoString PanCancer Immune Panel with PLUS codeset. Normalized counts were obtained using NanoString nSolver Software. Log2 transformed expression data were fit to a linear model comprised

of donor and treatment effects. Genes which showed a statistically significant treatment effect were identified in 3 initial donors. Adjusted p-values were used to correct for multiple hypothesis testing using the Benjamini-Hochberg procedure.

SIGNATURE CORRELATES WITH TUMOR RESPONSE



PRIOR TREATMENTS IN ADENOSINE SIGNATURE HIGH PATIENTS WITH PROLONGED PFS



Adenosine Signature levels were determined in pre-treatment biopsy tissue. In brief, RNA was extracted from tumor tissue macrodissected from patient biopsy specimens and analyzed using the NanoString PanCancer Immune Panel. Cutoffs for determining Adenosine Signature high and low tumors were based on the mean of the Log2 NanoString counts of select Adenosine Signature genes for all subjects evaluated. The t-test for comparing the maximum percentage decrease in SLD between the adenosine positive and negative signatures is 2-sided p-value = 0.0085, using a 5% level two-sided test.

AMP INDUCES ADENOSINE-LIKE GENE SIGNATURE

AMP INDUCED GENE SIGNATURE IS NEARLY IDENTICAL TO ADENOSINE SIGNATURE AMPas is a non-hydrolizable form of AMP - it does not form adenosine



CD73 ANTAGONISTS AMPLIFY AMP SIGNATURE

a NanoString Sprint instrument using the NanoString PanCancer Immune Panel with PLUS codeset. Normalized counts were obtained using NanoString nSolver Software. Representative data from one of >3 patients is shown above.

CD73 ANTAGONISTS INHIBIT ADENOSINE FORMATION, BUT CONSEQUENTLY PRESERVE AMP PRESERVED AMP AMPLIFIES ADENOSINE-LIKE GENE SIGNATURE SUGGESTING AMP SIGNALS THROUGH A2AR



PBS Control

CPI-006 AND CPX-016 ARE ANTI-CD73 ANTIBODIES. CP-1663 IS A CD73 SMALL MOLECULE INHIBITOR

Purified human PBMCs from healthy donors were co-cultured with CD73 antagonists including CPI-006 (Corvus anti-CD73 mAb, 10 μg/mL), CPX-016 (competitor anti-CD73 mAb, 10 μg/mL), an isotype control antibody, or CP-1663 (1 μM)) for 1 hour before adding AMP (150 μM). PBMCs were then stimulated after 1 hour with anti-human CD3 and CD28 antibodies. RNA was purified from cells collected after incubation for 48 hours. Gene expression changes were evaluated using the NanoString PanCancer Immune Panel. Representative data from one of >3 patients is shown above.



AMP IS AN ADENOSINE RECEPTOR AGONIST

AMP AND AMP αS AGONIZE A2AR, BUT ARE WEAKER THAN ADENOSINE

A2AR	EC50 (nM)	n
NECA	0.17	n = 2
ADENOSINE	2.7	n = 4
AMP	235.2	n = 4
AMPaS	228.6	n = 4
A1	EC50 (nM)	n
NECA	2.8	n = 2
ADENOSINE	11.9	n = 4
AMP	237.5	n = 4
AMPaS	418.2	n = 4

		1
A2BR	EC50 (nM)	n
NECA	1.3	n = 2
ADENOSINE	10.8	n = 4
AMP	227.5	n = 4
AMPaS	427.3	n = 4
A3	EC50 (nM)	n
NECA	1.7	n = 2
ADENOSINE	12.9	n = 4
AMP	199.0	n = 4
AMPaS	240.1	n = 4

cAMP assays were performed at Pharmaron using PerkinElmer CHO-K1 or HEK293 cells designed to stably overexpress human adenosine receptors, including hA1, hA2AR, hA2BR, and hA3. 3-fold dilutions of test compounds were evaluated, starting at a maximum concentration of 1 μ M (NECA) or 10 μ M (adenosine, AMP, or AMP α S). EC50 concentrations are shown along with the number of experiment replicates.

CIFORADENANT NEUTRALIZES AMP & AMPaS

CIFORADENANT BLOCKS AMP AND AMP as SIGNALING AT A2AR

A2AR	IC50 (nM)	n
ADENOSINE	4.0	n = 4
AMP	4.7	n = 2
AMPaS	7.1	n = 2

A1	IC50 (nM)	n
ADENOSINE	200.9	n = 4
AMP	111.4	n = 2
AMPaS	62.0	n = 2

ADENOSINE 171.2 $n = 4$ ΔMP 204.8 $n = 2$	A2BR	IC50 (nM)	n
ΔMP 204.8 n = 2	ADENOSINE	171.2	n = 4
	AMP	204.8	n = 2
AMPαS 270.2 n = 2	AMPaS	270.2	n = 2

A3	IC50 (nM)	n
ADENOSINE	516.0	n = 4
AMP	231.7	n = 2
AMPαS	315.1	n = 2

cAMP assays were performed at Pharmaron using PerkinElmer CHO-K1 or HEK293 cells designed to stably overexpress human adenosine receptors, including hA1, hA2AR, hA2BR, and hA3. Cells were stimultated with EC80 concentrations c indicated test compounds. Ciforadenant was added at a maximum concentration of 10 µM. IC50 values for ciforadenant inhibition of test compounds are shown, along with the number of experiment replicates.

CONCLUSIONS

 A2AR agonists induce a specific gene signature in human immune cells. This "Adenosine Signature" is dominated by inflammatory myeloid cytokines and chemokines.

Updated clinical data confirms original reports that expression of the Adenosine Signature correlates with tumor regression in an ongoing Ph 1/1b trial in patients with advanced/refractory RCC.

• High Adenosine Signature expression has a statistically significant correlation with tumor response.

The Adenosine Signature may be used as a predictive biomarker to select patients most likely to respond to therapy
with agents that antagonize adenosine production or signaling.

• AMP induces gene expression changes nearly identical to the Adenosine Signature in human immune cells, suggesting AMP can activate and signal through adenosine receptors. CD73 antagonists further amplify the AMP-associated gene expression changes as a consequence of preserving AMP.

• AMP is an agonist of adenosine receptors. AMP agonism of A2AR and A2BR can be blocked by ciforadenant.

 These data suggest treatment with CD73 antagonists will benefit from concominant blockade of A2AR to neutralize the resultant induction of AMP mediated gene expression changes.